The Value of Chromohysteroscopy in the Assessment of Postmenopausal Vaginal Bleeding

Yahia M. El-Faissal, Ahmed M. Kamel

Abstract

Background: To assess the endometrial changes in postmenopausal patients with vaginal bleeding, using chromohysteroscopy and histopathology, concluding consequently the value of chromohysteroscopy in investigating those patients.

Methods: We included a group of 50 postmenopausal women, presenting with vaginal bleeding, to the outpatient clinic of Cairo University hospital. Transvaginal 2D ultrasonography was performed, followed by hysteroscopy, with chromohysteroscopy performed. Hysteroscopic guided biopsies were obtained from stained areas, then formal curettage done to all uterine walls.

Results: Chromohysteroscopy patterns were either no staining (14%), staining of focal area(s) (76%) or staining of the whole endometrium (10%). Statistical significance was detected between endometrial thickness and chromohysteroscopy. We found a sensitivity 93.75%, specificity 27.77%, PPV 69.76%, and NPV 71.42% of chromohysteroscopy for the pathology obtained.

Conclusion: Chromohysteroscopy improves the efficacy of hysteroscopy in targeting endometrial biopsy. It is though labor intensive and yet not standardized. Studies involving different dyes and larger number of participants are warranted. Inter-observer variability is equally important to help standardizing the procedure.

Keywords: Chromohysteroscopy; Postmenopausal bleeding; Endometrial change

Introduction

Postmenopausal bleeding is a common gynecologic complaint, representing up to 69% of postmenopausal women referred to gynecological clinics [1]. Approximately 90% of women with endometrial carcinoma report vaginal bleeding as the sole complaint, so this symptom should always be carefully investigated. However, just 10-15% of women with postmenopausal bleeding have endometrial carcinoma [2].

Dilatation and curettage (D&C) was, for long, considered the “gold standard” for the investigation of postmenopausal bleeding [3]. As a blind procedure, it often results in unrepresentative biopsies [4] with a diagnostic failure that varies from 10 to 25% [5], and false negative rates between 2 and 10%. Older reports have revealed that in 60% of women submitted to curettage less than half of the uterine cavity was sampled with the curette and that the source of bleeding was frequently not diagnosed [6].

Diagnostic hysteroscopy has now replaced conventional cervical D&C for the evaluation of the uterine cavity in industrialized countries. Diagnostic hysteroscopy can be regarded as the gold standard for the evaluation of the uterine cavity. Hysteroscopic surgery is a safe and cost-effective diagnostic and therapeutic tool for the investigation and treatment of intrauterine anomalies [7].

Transvaginal ultrasonography with high-resolution transducers allows a good definition of the endometrial-myometrial interface and visualization of the entire endometrial cavity. Transvaginal ultrasonography is a highly sensitive test for detecting endometrial disease, but carries a false negative rate of 8% for endometrial carcinoma. Several reports showed that the combined assessment of endometrial thickness with some morphologic parameters improves the diagnostic accuracy of transvaginal sonography in patients with postmenopausal bleeding [8].

Chromoendoscopy is a widely used technique in gastrointestinal imaging [9]. Over the last decade, endoscopic systems have acquired great power due to high-resolution images owing to charge-coupled device chip technology and narrow band imaging technique [10]. Based on this growing and advanced applications, chromohysteroscopy was first introduced after the study by Kucuk and Safali in 2008, where they combined chromoendoscopy and hysteroscopy as a new avenue to improve the diagnostic value of hysteroscopy in the setting of assisted reproduction. In this work, we studied...
the application of chromohysteroscopy in the diagnosis of postmenopausal uterine bleeding [11].

**Aim of work**

The aim of this study is to correlate the changes occurring in the endometrium of postmenopausal patients with vaginal bleeding, using chromohysteroscopy, together with histopathological examination of biopsies taken from that endometrium. Consequently, and upon analyzing such data, we would reach a conclusion concerning the value of adding chromohysteroscopy as a tool to investigate a patient with postmenopausal vaginal bleeding, as well as its actual impact on the diagnosis.

**Materials and Methods**

A group of consecutive 50 postmenopausal women (no menstrual periods for 12 consecutive months and no other biological nor physiological cause could be identified), presenting with vaginal bleeding, to the outpatient clinic of Cairo University Hospital (Kasr El Eini University Hospital) were included in this study from January 2012 till September 2012. The study has been approved by the internal review board and the Ethical Committee of the Faculty of Medicine, Cairo University.

The patients were properly assessed according to the department’s protocol; for those who were included in the study, a written informed consent of the hysteroscopy procedure was signed by each, including verbal explanation addressing all the details of the procedure of chromohysteroscopy. Transvaginal two-dimensional (2D) ultrasonography, using a GE Voluson E8 ultrasound system (GE Healthcare Ultrasound, Milwaukee, WI, USA) equipped with a 6 - 12 MHz 3D endovaginal probe, was then performed for each, including visual evaluation of uterine dimensions, endometrial thickness and pattern (homogeneity and echogenicity),

**Table 1. Age Distribution and Parity of Patients Included in the Study**

<table>
<thead>
<tr>
<th>Number of</th>
<th>Mean</th>
<th>SD</th>
<th>Minimum</th>
<th>Maximum</th>
<th>Median</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age distribution (in years)</td>
<td>50</td>
<td>57.5</td>
<td>6.3</td>
<td>47</td>
<td>72</td>
</tr>
<tr>
<td>Parity</td>
<td>50</td>
<td>3.4</td>
<td>1.3</td>
<td>1</td>
<td>6</td>
</tr>
</tbody>
</table>

**Table 2. The Correlation Between Endometrial Thickness, Histopathological Diagnosis and Pattern of Subendometrial Perfusion Obtained by Power Doppler Endometrial**

<table>
<thead>
<tr>
<th>Number</th>
<th>Mean</th>
<th>Std deviation</th>
<th>Median</th>
<th>Min.</th>
<th>Max.</th>
<th>Kruskal-Wallis test</th>
</tr>
</thead>
<tbody>
<tr>
<td>Atrophy</td>
<td>N=14</td>
<td>4.4</td>
<td>1.2</td>
<td>4.0</td>
<td>3.0</td>
<td>8.0</td>
</tr>
<tr>
<td>Endometritis</td>
<td>N=5</td>
<td>8.0</td>
<td>3.7</td>
<td>6.0</td>
<td>5.0</td>
<td>12.0</td>
</tr>
<tr>
<td>Endometrial hyperplasia</td>
<td>N=4</td>
<td>16.5</td>
<td>4.7</td>
<td>15.5</td>
<td>12.0</td>
<td>23.0</td>
</tr>
<tr>
<td>Endometrial cancer</td>
<td>N=4</td>
<td>25.3</td>
<td>5.1</td>
<td>25.0</td>
<td>20.0</td>
<td>31.0</td>
</tr>
<tr>
<td>NEP</td>
<td>N=23</td>
<td>8.0</td>
<td>2.2</td>
<td>7.0</td>
<td>5.0</td>
<td>13.0</td>
</tr>
</tbody>
</table>

*: Statistically significant. NEP: No endometrial pathology.
regularity of the endometrial-myometrial border and fluid in
the cavity.

These investigations were done to the patients at the
time of their primary visit. They were then scheduled for
diagnostic hysteroscopy, during which chromohysteroscopy
was performed. A TROPHYSCOPE™ - CAMPO Compact
Hysteroscope (Karl Storz®, Tuttingen, Germany), with con-
tinuous flow sheath, 5 Fr working channel was used. Dis-
tilled water (H2O) was used for uterine distension, with an
insufflator that maintained a 100 - 150 mmHg pressure in the
uterine cavity. Cervical dilatation was not needed in most of
the cases. Those who needed cervical dilatation, were sched-
uled for diagnostic hysteroscopy under general anesthesia,
during which cervical dilatation was performed.

During the setting for chromohysteroscopy, 5 - 10 mL
of 2% solution of "methylene blue" (methylthioninium
chloride; C16H18N3SCl) (Muby Chemicals, Chinchbunder,
Mumbai, India) were instilled into the uterine cavity using
a disposable sterile 20 mL plastic syringe connected to the
inflow port of the hysteroscope. Uterine distension with dis-
tilled water was then resumed for one whole minute, in order
to distribute and flush the dye. The patterns of staining of
the endometrium were then noted, and were allocated, for
statistical convenience, to be one of either:

1) No staining: When no part of the endometrium retains
the dye upon examining with the hysteroscope for at least 5
min.

2) Staining of focal area(s) of the endometrium: When a
single or multiple localized areas of the endometrium appear
more darkly stained than the remainder of the endometrium,
following examining with the hysteroscope for at least 5 min.

3) Staining of the whole endometrium: When the entire
surface of the endometrium appears homogenously stained
with the dye following examining with the hysteroscope for
at least 5 min.

Tissue samples were then obtained from stained areas
under hysteroscopic guidance. In cases where there is no
staining of the endometrium, random endometrial biopsies
were taken from the anterior and posterior walls of the uter-
us, as well as the uterine fundus.

The biopsies obtained were sent to the Pathology De-
partment of Cairo University, for routine histopathological
examination. The results were recorded, and then compared
with, and correlated to the hysteroscopy findings.

According to the findings, each patient was properly
managed according to the institute’s policy.

Statistical methods

Data were analyzed using SPSSwin statistical package ver-
sion 15 (IBM, Armonk, NY, USA). Numerical data were
expressed as mean and standard deviation or median and
range as appropriate. Qualitative data were expressed as fre-
quency and percentage. Chi-square test (Fisher’s exact test)
was used to examine the relation between qualitative vari-
ables. For quantitative data, comparison between two groups
was done using Mann-Whitney test (non-parametric t-test).
Comparison between more than two groups was done using
Kruskal-Wallis test (non-parametric ANOVA), and then post
hoc “Schefe test” on rank of variables was used for pair-wise
comparison. Pearson’s correlation was used to test relation
between numerical variables. A P value < 0.05 was consid-
ered significant.

Results and Discussion

The patients’ age ranged from 42 to 72 years, with an aver-
age mean of 57.5 years. While the parity ranged from 1 to 6
with an average of 3.4 times (Table 1).

Endometrial thickness in this study was estimated by
performing the measurement between the two basal layers of
the anterior and posterior uterine walls. The poorly reflective
layer surrounding the highly reflective endometrium was not
included in the measurement. When the endometrial layers
were separated with fluid, both layers were measured and the
sum was recorded. This is in accordance with the method of measurement adopted in previous studies such as that of Van Den Bosch et al [12] and Sousa et al [2]. However endometrial thickness has been measured in different ways by other authors, such as Fleischer et al [13] who measured the cavity whole through and divided this into two as a measure of single layer endometrial thickness. Nasri and Coast [14] estimated the measure of the endometrium single layer and excluded distension of fluid. Smith et al [15] also recorded a single layer measurement. The endometrial thickness data are shown in Table 2. It is to be noted that the mean values, as well as the distribution of individual measurements, could not yield a cut-off limit for endometrial thickness to identify the pathological endometrium from the normal. This fact is explained by observing the ranges of thickness; the smallest endometrial thickness was 3.0 mm. The difference in endometrial thickness in relation to pathology obtained through chromohysteroscopy revealed a mean of 4.2 mm with a minimum of 3.0 mm in cases with atrophy, a range between 5.0 and 12 mm in endometritis; a mean of 16.5 mm in endometrial hyperplasia with a maximum of 23.0 mm. Cases with endometrial cancer showed a mean endometrial thickness of 25.3 mm, while those with bleeding and non-significant endometrial pathology had a range between 5.0

<table>
<thead>
<tr>
<th>Chromohysteroscopy pathology</th>
<th>Focal stain</th>
<th>Whole end</th>
<th>No stain</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Atrophy</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Count</td>
<td>12</td>
<td>0</td>
<td>2</td>
<td>14</td>
</tr>
<tr>
<td>% within chromohyst pathology</td>
<td>85.7%</td>
<td>0%</td>
<td>14.3%</td>
<td>100.0%</td>
</tr>
<tr>
<td>% within chromohys stain</td>
<td>31.6%</td>
<td>0%</td>
<td>28.6%</td>
<td>28.0%</td>
</tr>
<tr>
<td><strong>Endometritis</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Count</td>
<td>5</td>
<td>3</td>
<td>0</td>
<td>8</td>
</tr>
<tr>
<td>% within chromohyst pathology</td>
<td>62.5%</td>
<td>37.5%</td>
<td>0%</td>
<td>100.0%</td>
</tr>
<tr>
<td>% within chromohys stain</td>
<td>13.2%</td>
<td>60.0%</td>
<td>0%</td>
<td>16.0%</td>
</tr>
<tr>
<td><strong>Endo hyperplasia</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Count</td>
<td>6</td>
<td>0</td>
<td>0</td>
<td>6</td>
</tr>
<tr>
<td>% within chromohyst pathology</td>
<td>100.0%</td>
<td>0.0%</td>
<td>0.0%</td>
<td>100.0%</td>
</tr>
<tr>
<td>% within chromohys stain</td>
<td>15.8%</td>
<td>0.0%</td>
<td>0.0%</td>
<td>12.0%</td>
</tr>
<tr>
<td><strong>Endo cancer</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Count</td>
<td>2</td>
<td>2</td>
<td>0</td>
<td>4</td>
</tr>
<tr>
<td>% within chromohyst pathology</td>
<td>50.0%</td>
<td>50.0%</td>
<td>0.0%</td>
<td>100.0%</td>
</tr>
<tr>
<td>% within chromohys stain</td>
<td>5.3%</td>
<td>540.0%</td>
<td>0.0%</td>
<td>8.0%</td>
</tr>
<tr>
<td><strong>NEP</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Count</td>
<td>13</td>
<td>0</td>
<td>5</td>
<td>18</td>
</tr>
<tr>
<td>% within chromohyst pathology</td>
<td>72.2%</td>
<td>0.0%</td>
<td>27.8%</td>
<td>100.0%</td>
</tr>
<tr>
<td>% within chromohys stain</td>
<td>34.2%</td>
<td>0.0%</td>
<td>71.4%</td>
<td>36.0%</td>
</tr>
<tr>
<td><strong>Total</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Count</td>
<td>38</td>
<td>5</td>
<td>7</td>
<td>50</td>
</tr>
<tr>
<td>% within chromohyst pathology</td>
<td>76.0%</td>
<td>10.0%</td>
<td>14.0%</td>
<td>100.0%</td>
</tr>
<tr>
<td>% within chromohys stain</td>
<td>100.0%</td>
<td>100.0%</td>
<td>100.0%</td>
<td>100.0%</td>
</tr>
</tbody>
</table>

NEP: No endometrial pathology.

Table 3. Different Chromohysteroscopy Patterns With Variable Pathology Results
and 13.0 mm. It should be stated, however, that based on these results, the estimation of endometrial thickness in postmenopausal women could provide a good screening method, helping to predict those women who are liable to experience postmenopausal bleeding.

When using the Kruskal-Wallis test to compare the pathology obtained from either fractional curettage or chromohysteroscopy to the endometrial thickness, as shown in Table 2, a P value of < 0.001 was found giving a positive statistical significance.

Chromohysteroscopy

As chromohysteroscopy is a new novel technique in obtaining a targeted endometrial biopsy, the stain used and the method of instillation of the dye and data interpretation were adapted from the study by Kucuk and Deveci [16]. The staining pattern was divided as previously described.

The data gathered showed 76% with focal staining, 14% with no stain and 10% with the whole endometrium stained (Fig. 1). The samples obtained were sent for routine histopathological examination which revealed atrophy in 28%, endometritis in 16%, endometrial hyperplasia in 12%, cancer in 8% and no endometrial pathology (NEP) in 36% of cases.

Staining pattern revealed that out of 14 cases of atrophy, 12 showed a focal staining pattern and in the remaining two cases, there was no staining of the endometrium. However, endometritis showed five cases of focal staining and three cases with the whole endometrium stained. All cases of endometrial hyperplasia showed focal staining, while endometrial carcinoma showed focal staining in half of the cases and the other half showed staining of the whole endometrium. Cases with non-significant endometrial pathology showed a focal staining pattern in 13 out of the 18 cases, with the rest showing no staining pattern (Table 3). Chromohysteroscopy led to the diagnosis of three more cases of endometritis and two more cases of focal endometrial hyperplasia, compared to the standard blind fractional curettage. The difference in positive results between fractional curettage and chromohysteroscopy was statistically significant (Table 4).

Kucuk and Deveci found two extra cases of endometritis and one more case of endometrial hyperplasia when using chromohysteroscopy technique compared to non-targeted endometrial biopsy. Both the results of our study and their study showed greater accuracy in detecting pathological changes in the endometrium. The difference between the results of biopsy by chromohysteroscopy and fractional curettage was statistically significant in our study. They never commented on the statistical significance of their results, probably due to the small number of patients included in their study (22 patients). Another more recent study by Mansour and Mohamed [17] showed that diffuse light staining does not match well with histopathology, so it cannot be relied upon to prove or exclude chronic endometritis. On the other hand, focal dark staining showed sensitivity (70%), specificity (80.8%), positive predictive value (PPV) (43.7%) and negative predictive value (NPV) (92.6%). The result of focal dark staining matched well with the results of histopathology. They consequently concluded that focal dark staining can be used to prove or exclude chronic endometritis.

The sensitivity of chromohysteroscopy technique was 93.75%, specificity was 27.77%, PPV was 69.76% and NPV was 71.42%.

When calculating the sensitivity, specificity, positive and negative predictive values of chromohysteroscopy for the pathology obtained (where positive results = stain (focal

<table>
<thead>
<tr>
<th>Table 4. Comparison Between the Different Pathology Results Sampled Through Chromohysteroscopy and Ordinary Fractional Curettage</th>
</tr>
</thead>
<tbody>
<tr>
<td>Atrophy</td>
</tr>
<tr>
<td>-----------------</td>
</tr>
<tr>
<td>Fractional curettage (cases)</td>
</tr>
<tr>
<td>Chromohysteroscopy (cases)</td>
</tr>
</tbody>
</table>

*: Statistically significant.
and whole endometrium), negative results = no stain, true positive = atrophy, endometritis, endometrial hyperplasia and cancer … with stain, true negative = NEP with no stain, false positive = NEP with stain and false negative = atrophy, endometritis, endometrial hyperplasia and cancer … with no stain), we found a sensitivity of 93.75%, specificity of 27.77%, PPV of 69.76% and NPV of 71.42%. The low specificity recorded in our study is the result of the cases with NEP becoming stained during the procedure; our study did not take into account the varying degrees of stain depth, namely, we did not differentiate between dark and light staining of the endometrium as did the study by Mansour and Mohamed; however, they did not present any criteria identifying the degrees of staining nor the threshold at which the dyeing process was considered whether light or dark apart from the naked eye appreciation of the degree of color. We found such judgment non-standardized and poorly reproducible.

Conclusions and recommendations

In conclusion, chromohysteroscopy improves the efficacy of hysteroscopy in targeting endometrial biopsy. Our study implies endometrial staining with methylene blue dye improves the diagnostic value and provides a method for guided biopsy to diagnose endometrial pathology during hysteroscopy in the absence of macroscopic abnormalities. Chromohysteroscopy seems to be a candidate for implementation in surveillance programs for high risk patients, but it is a labor intensive and time consuming technique, which so far has prevented its wide spread use. Future studies involving chromohysteroscopy should focus on its value in clinical practice by average experienced endoscopists. Other studies should be carried out with different dyes, as different dyes have different staining properties, and some will be more suited for infection, while others more suited for dysplasia and neoplasia. More extensive research is still warranted in order to establish the previous suggestions. Studies that include a larger number of patients and long-term follow-up are required to verify the real predictive value of chromohysteroscopy in postmenopausal bleeding, as well as their role in the diagnosis. Lastly, it is of great importance to carry out further studies concerning the inter-observer and the intra-observer reproducibility, and with accumulating results, a standardized protocol could be established.

Conflicts of Interest

The authors have no conflicts of interest to disclose.

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target endometrial biopsy in postmenopausal bleeding?
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